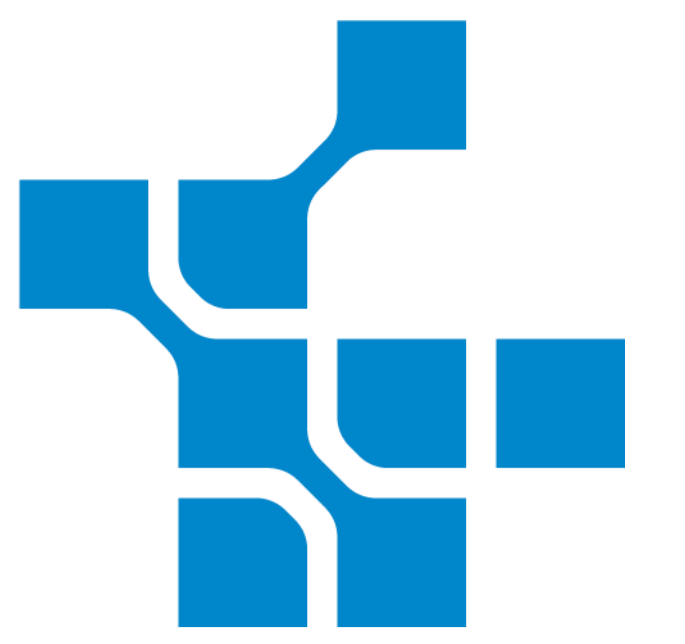


An innovative approach for conducting 3D electrophysiological recordings within intact brain organoids



Sara Mirsadeghi^{1,2}, Tom Stumpp³, Michael Mierzejewski³, Angelika Stumpf³, Haein Chang³, Udo Kraushaar³, Ali Hosseini⁴, Sven Schoenecker⁵, Julio Alvarez⁵, Michele Giugliano⁶, Jenny Hsieh^{1,2} and Peter D. Jones³

peter.jones@nmi.de

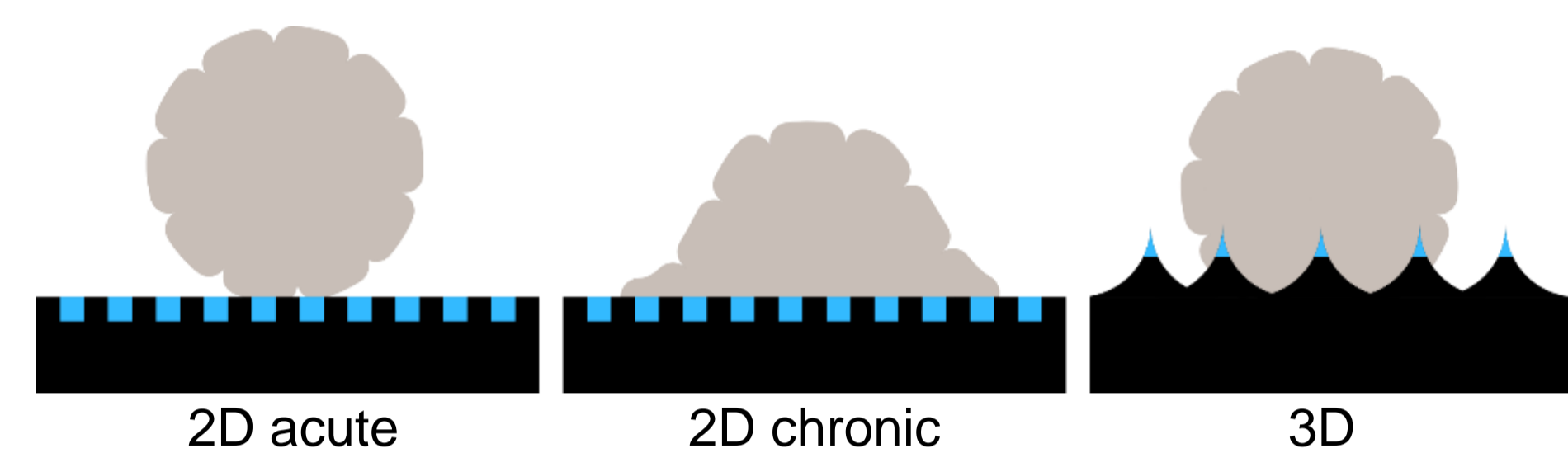
1. Department of Neuroscience, Developmental and Regenerative Biology, The University of Texas at San Antonio, San Antonio, TX, USA
2. Brain Health Consortium & Department of Neuroscience, Developmental and Regenerative Biology, The University of Texas at San Antonio, San Antonio, TX, USA
3. NMI Natural and Medical Sciences Institute at the University of Tübingen, 72770 Reutlingen, Germany

4. International School of Advanced Studies, Neuroscience Area, Trieste, Italy and Sorbonne Université, CNRS, ISIR, Paris, France
5. Multi Channel Systems MCS GmbH (a Harvard Bioscience company), Reutlingen, Germany
6. Bioengineering department, University of Modena and Reggio Emilia, Italy

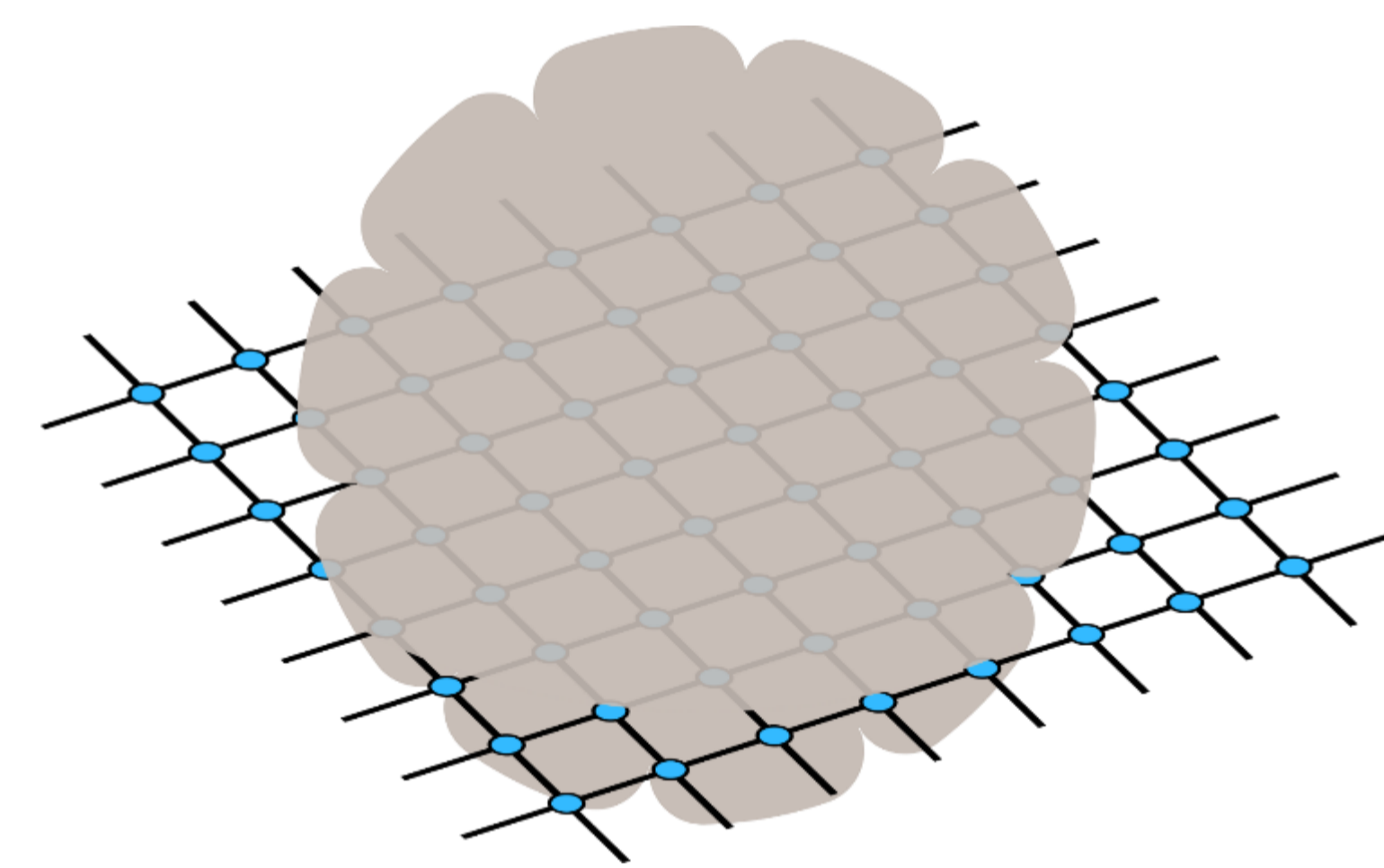
Introduction and summary

- Neurological disorders often lack translatability in animal models.¹
- iPSC-derived brain organoids offer a human-relevant model system.¹
- Electrophysiology is crucial for assessing neural circuit activity and drug effects.
- We present a mesh microelectrode array (MEA) device for 3D neural tissue *in vitro*.^{2,3}
- Compared to classical planar MEAs, the mesh MEA should allow chronic recordings and minimal disruption to the 3D tissue.
- Neural tissue envelops the mesh filaments, enabling recordings from within the tissue.
- The mesh enables medium flow from all sides and keeps the tissue suspended away from all surfaces.

Spheroids are constrained by the solid surfaces of planar or 3D MEAs, which may diminish physiological relevance.

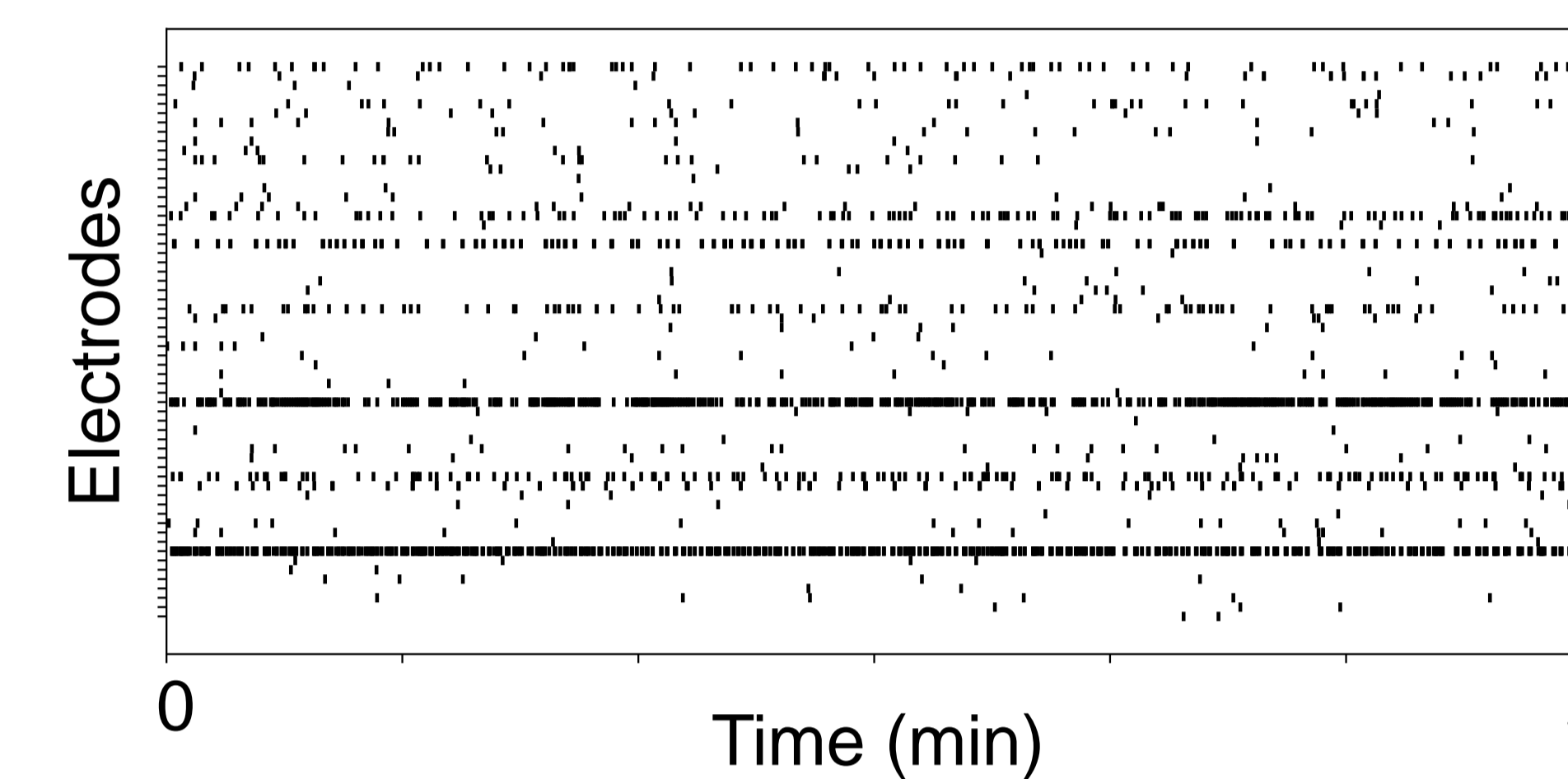
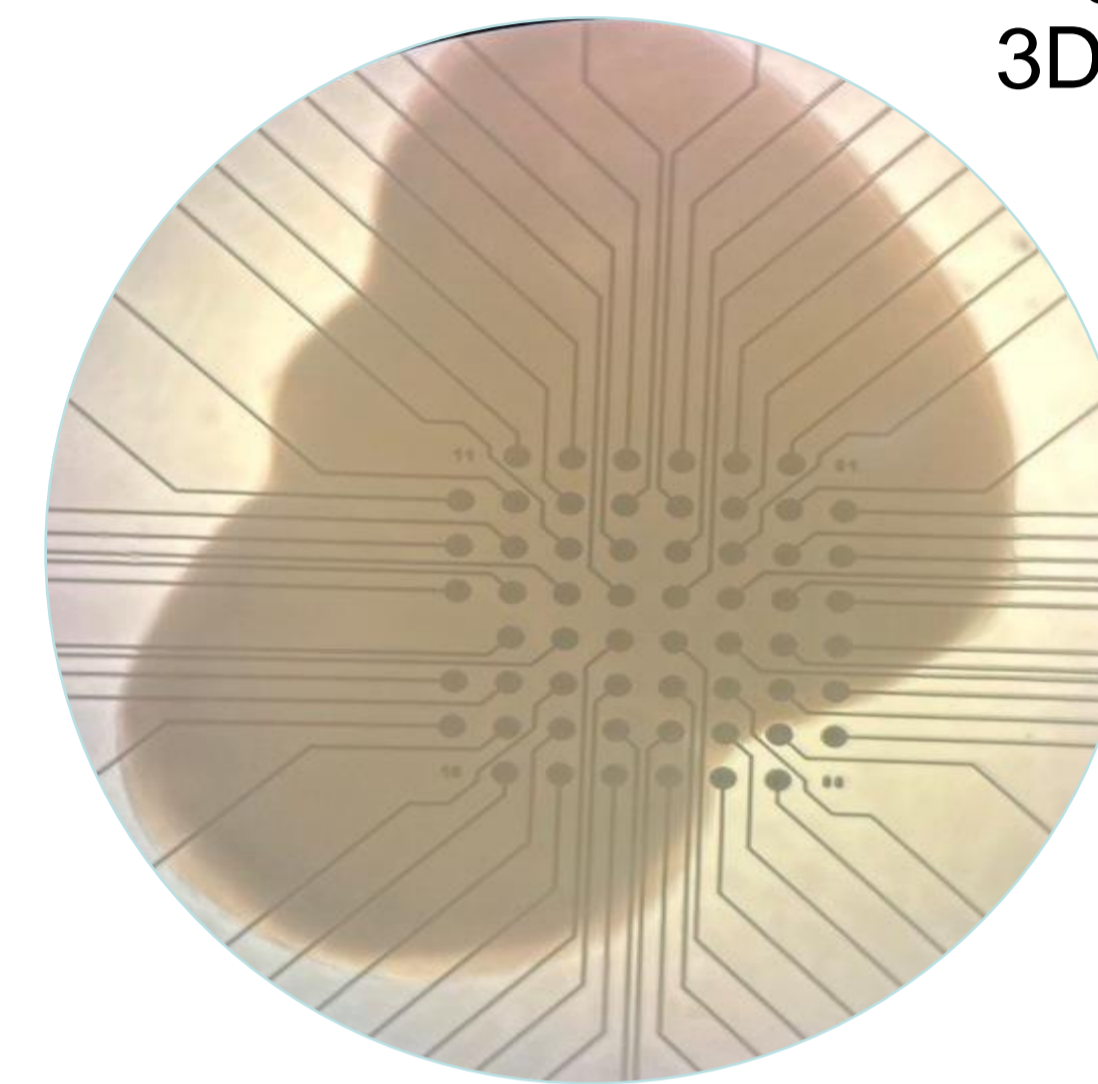


Mesh MEAs do not constrain spheroids, and microelectrodes are internalized to record from within 3D structures.

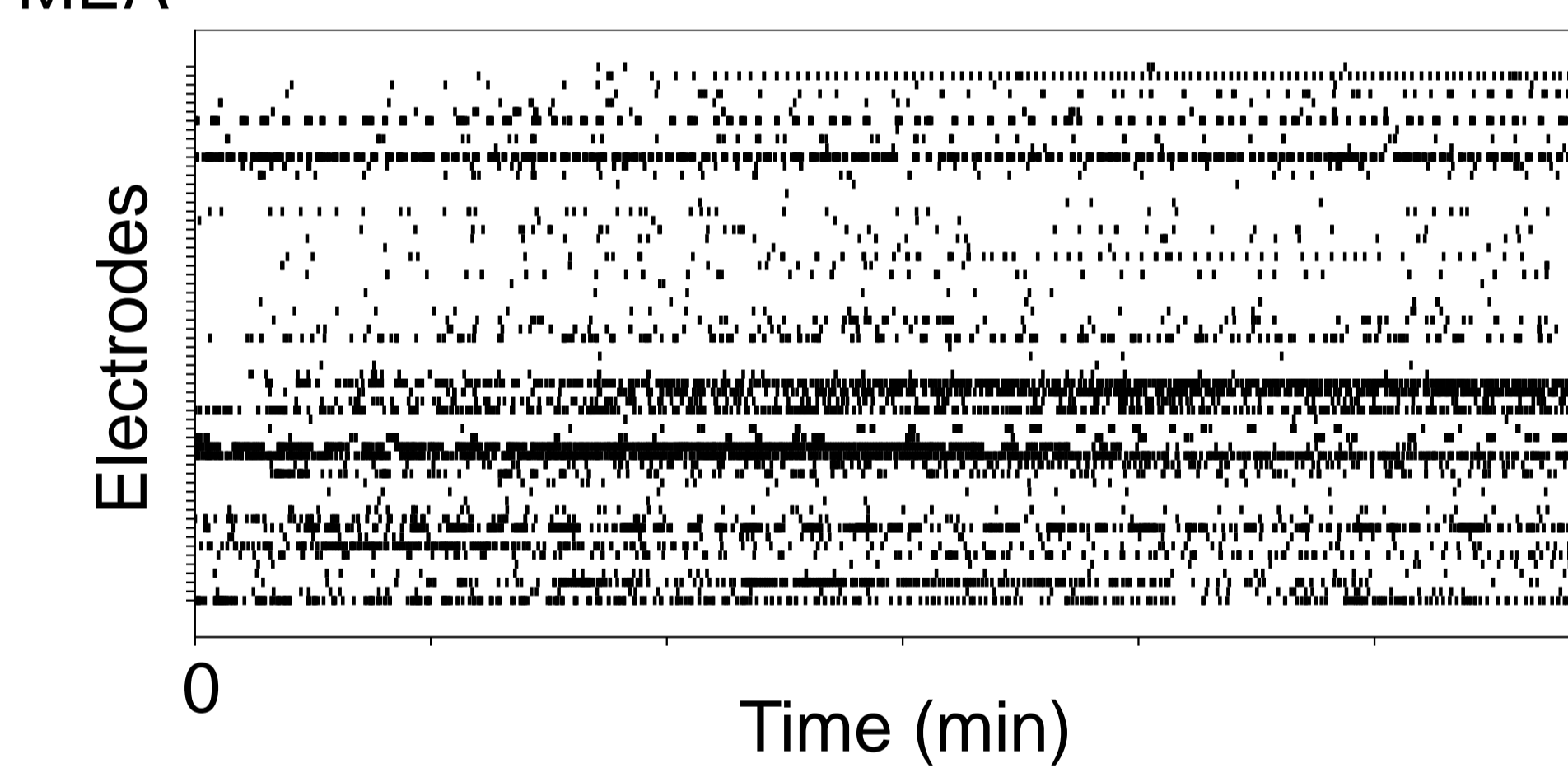
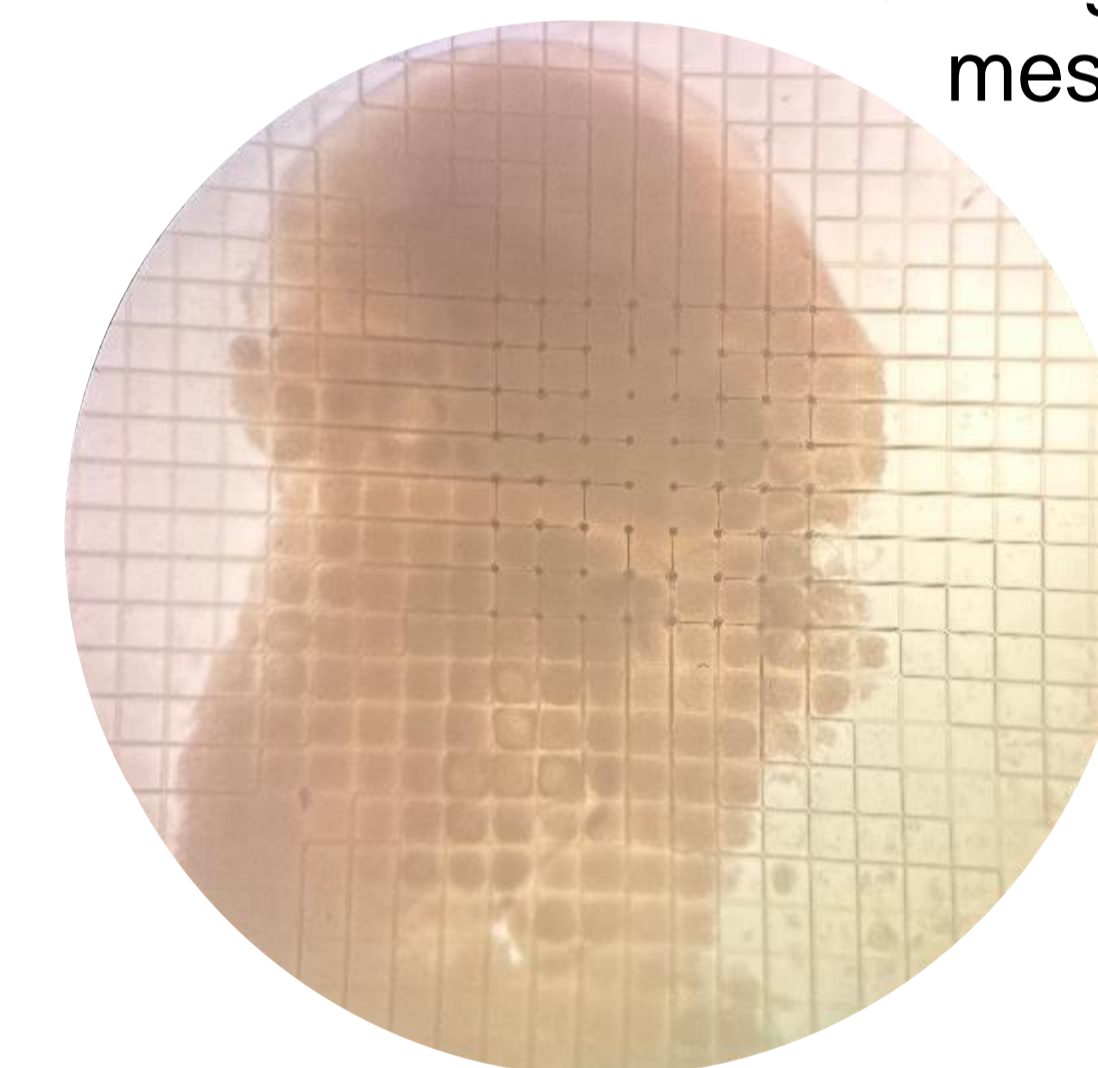


Recording of human brain organoids

Brain organoid on a 3D MEA



Brain organoid on a mesh MEA



Outlook

- Mesh MEAs allow repeated long-term measurements within neural spheroids, organoids, and other 3D tissue models.
- The well design allows simple medium exchange, culture at an air-liquid interface, and perfusion.
- This new tool should contribute to a better understanding of electrophysiological activity in 3D *in vitro* models.
- Functional electrical readout will help to develop advanced 3D models of human neurodevelopment and disease.

Contributions & acknowledgements

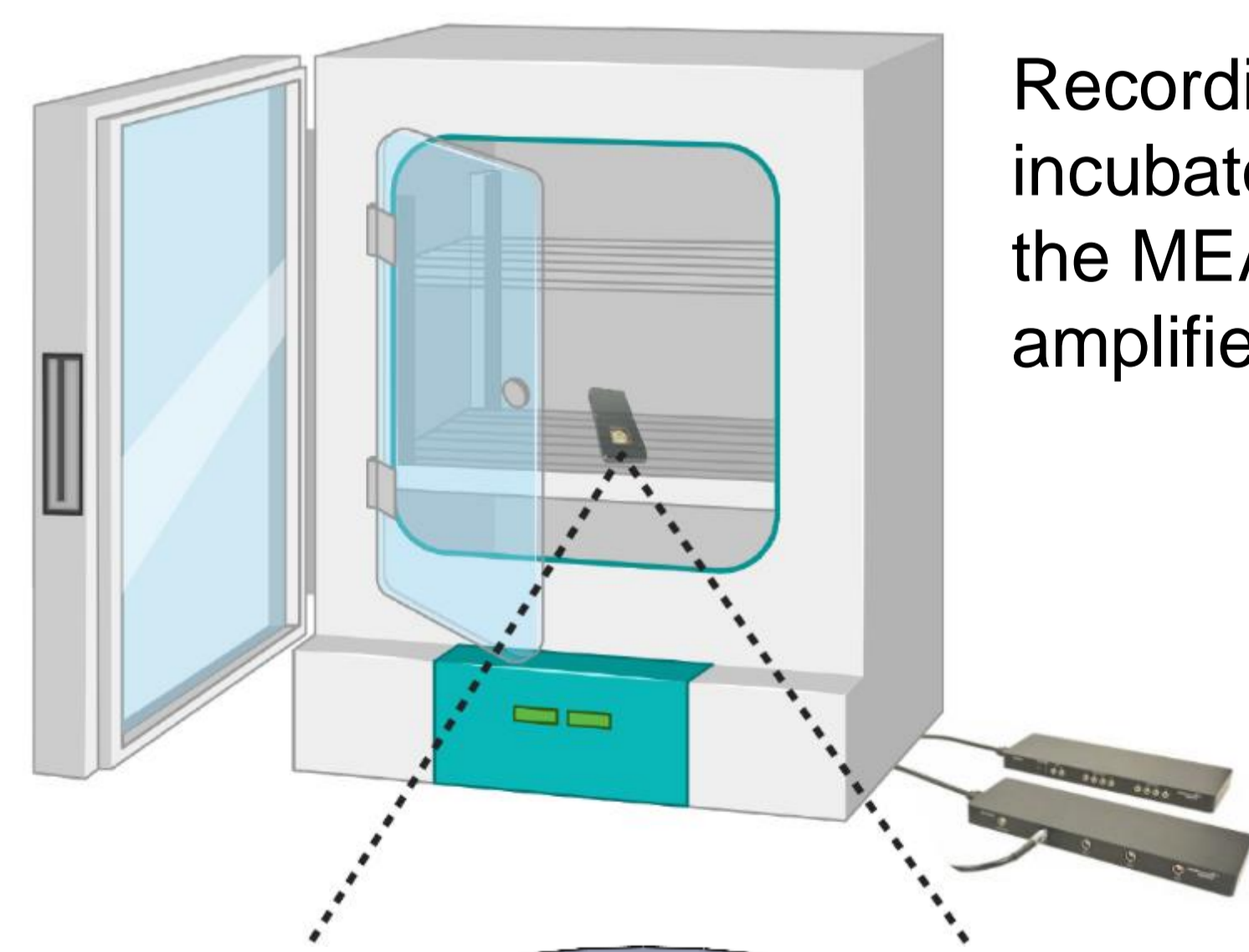
TS, HC, MM, DP, AS, HC & UK & PDJ: development and fabrication of mesh MEAs and evaluation using neural spheroids.
SM, JH: evaluation of MEAs using brain organoids
AH, MG: analysis of recordings
SS, JA: amplifier hardware and software

NMI acknowledges support from the German Research Foundation (DFG) in project MEMMEA (#441918103) as part of the DFG priority program SPP 2262 MemrisTec (#422738993) and from the State Ministry of Baden-Wuerttemberg for Economic Affairs, Labour and Tourism. UTSA acknowledges NIH grants U01DA054170, R01NS113516, and R01NS124855 and funding from the Robert J. Kleberg, Jr. and Helen C. Kleberg Foundation and the Semmes Foundation.

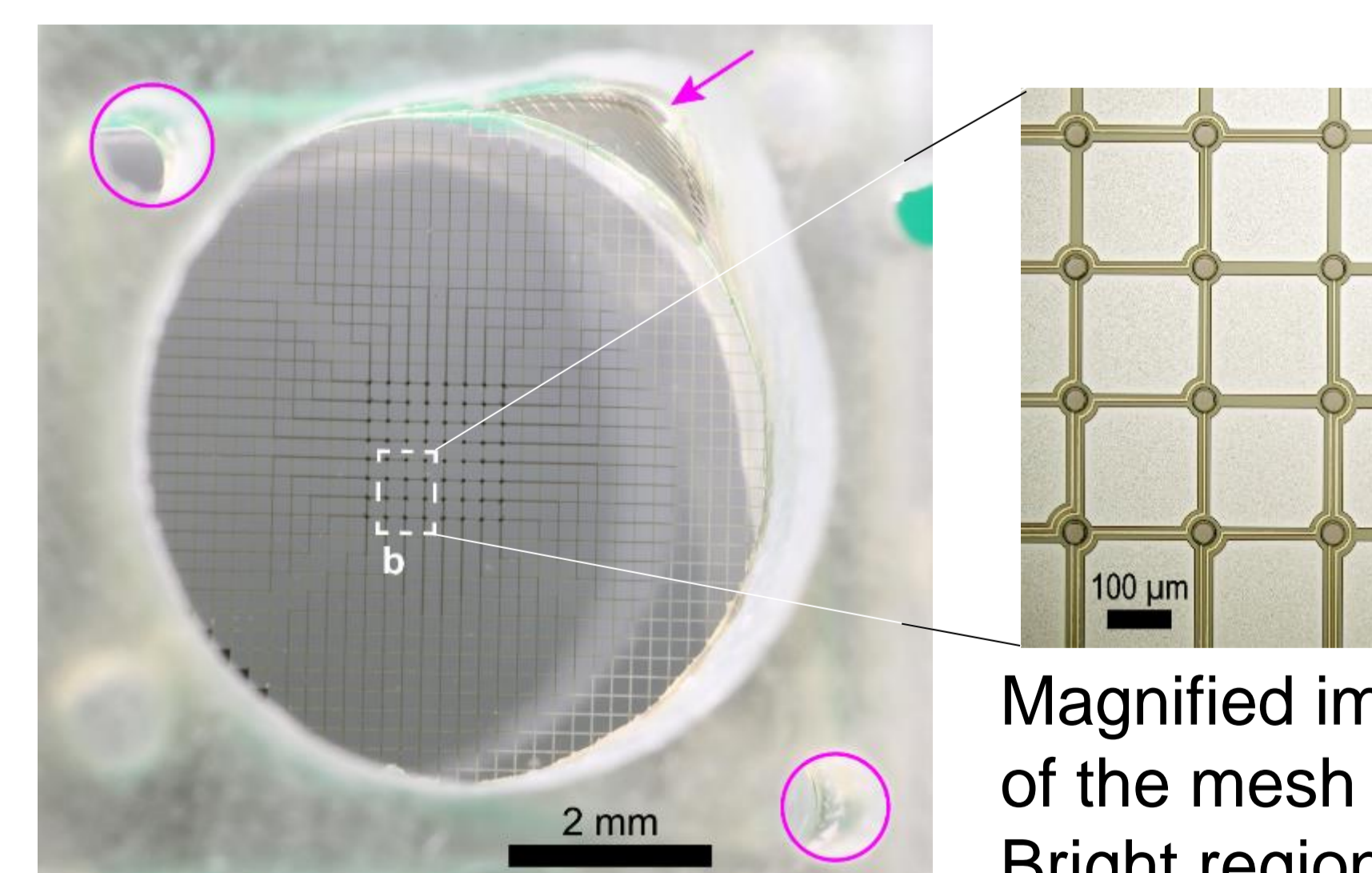
To learn more about using mesh MEAs in your lab, visit the Harvard Bioscience booth.



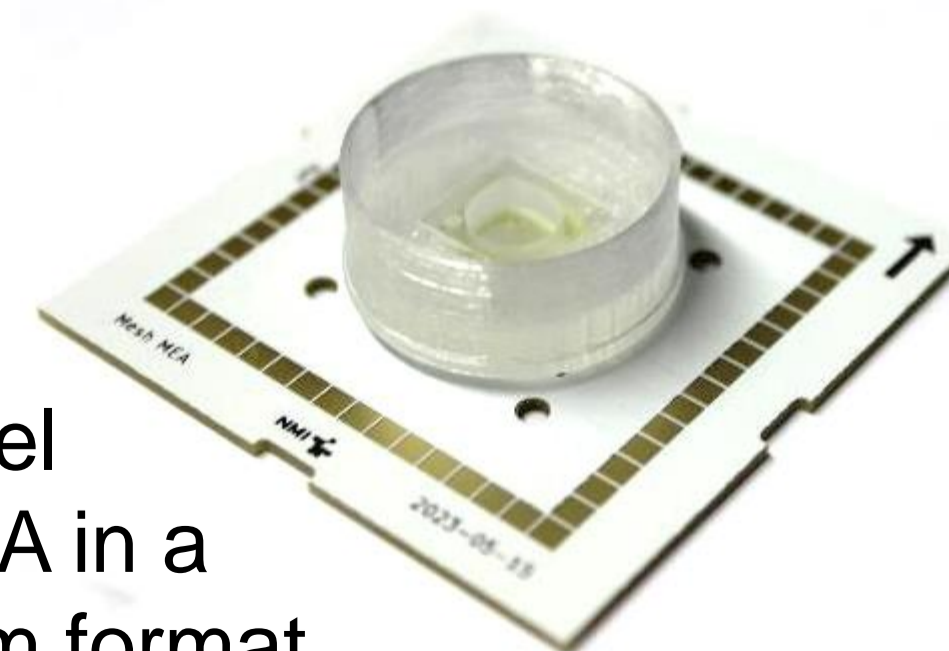
Mesh MEA design



Recordings in an incubator using the MEA Mini amplifier (MCS).



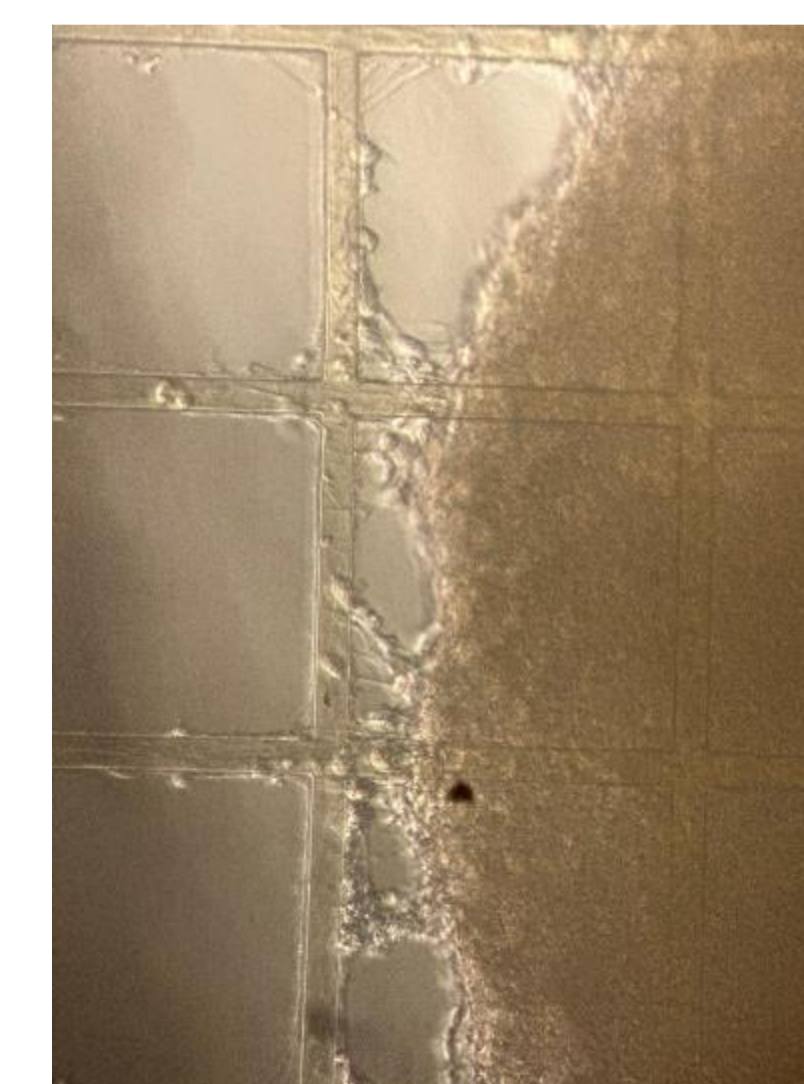
Inner well with two in-/outlets and a pipetting ledge. Magnified image of the mesh MEA. Bright regions are open space.



60-channel mesh MEA in a 49x49 mm format.

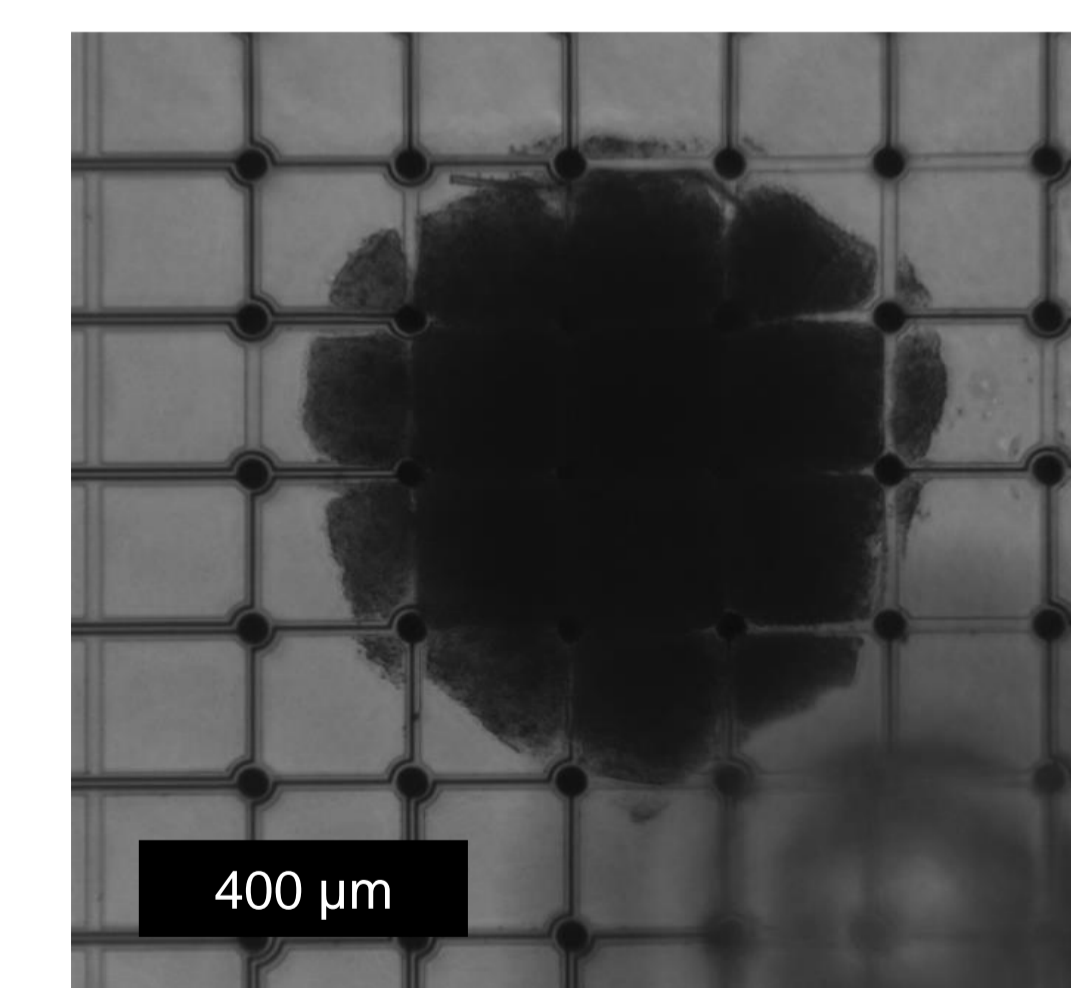
Cross section of the mesh MEA. The mesh suspends spheroids 2 mm above the bottom window.

Neuron growth on the mesh

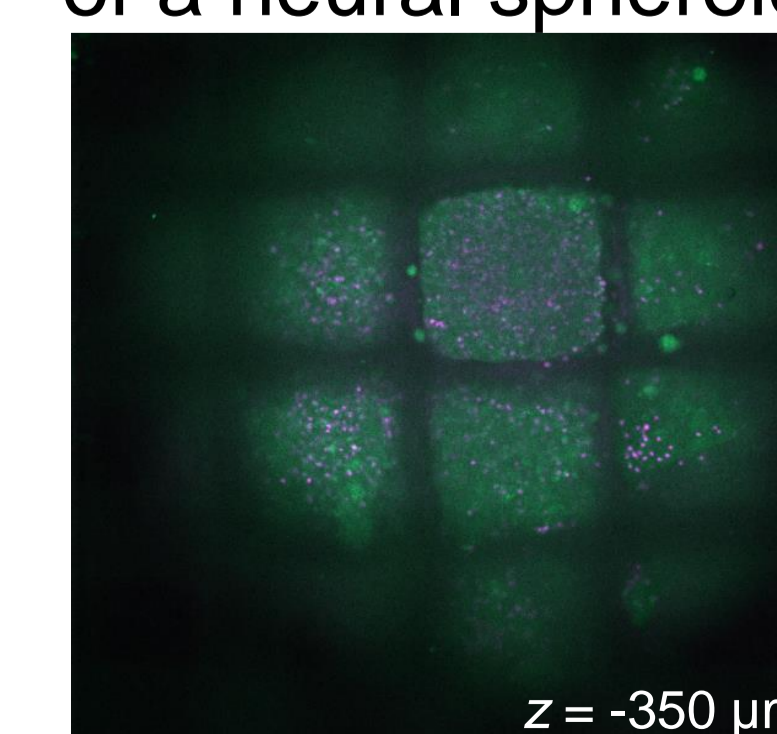
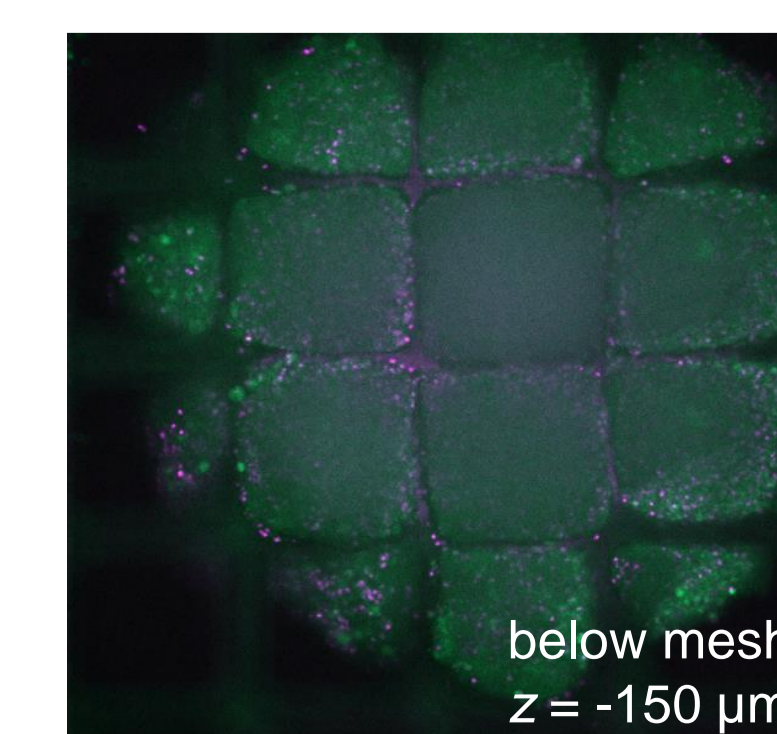
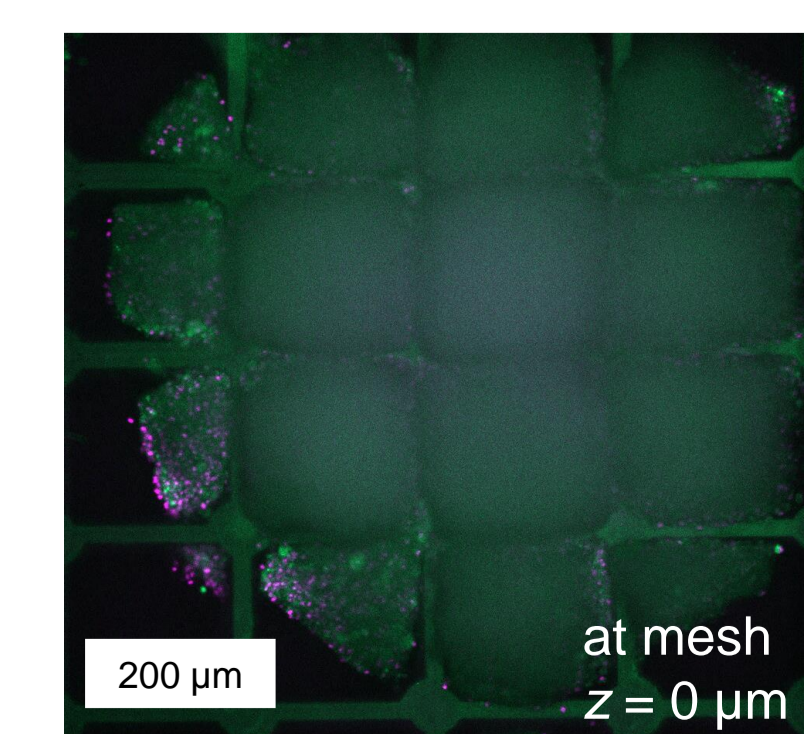


Neuronal migration after seeding shows cell viability

Neural spheroid placed on the mesh



Live and dead staining of a neural spheroid



Propidium iodide (dead)
Cal-520AM (alive)

References

- [1] Engle, S.J., Blaha, L., Kleiman, R.J., (2018) Best Practices for Translational Disease Modeling Using Human iPSC-Derived Neurons. *Neuron*, 100, 783–797.
- [2] Stumpp, T.; Mierzejewski, M.; Pascual, D.; Stumpf, A.; Jones, P. D. (2023) Scalable Mesh Microelectrode Arrays for Neural Spheroids and Organoids. *Current Directions in Biomedical Engineering*, 9 (1), 575–578.
- [3] McDonald, M. *et al.* (2023) A Mesh Microelectrode Array for Non-Invasive Electrophysiology within Neural Organoids. *Biosensors and Bioelectronics*, 228,